

# Information design and data visualization

## Week 3, Lecture 06

*Richard E.W. Berl*

*Spring 2019*

### ggplot2

```
library(ggplot2)
```

Load the egg data as a CSV.

```
egg = read.csv("../data/aaj1945_DataS1_Egg_shape_by_species_v2.csv", header=T,  
              stringsAsFactors=F)
```

```
colnames(egg)[7:9] = c("AvgLength", "NumberOfImages", "NumberOfEggs")
```

```
egg = egg[-c(1401,1402),]
```

```
str(egg)
```

```
## 'data.frame': 1400 obs. of 9 variables:  
## $ Order : chr "ACCIPITRIFORMES" "ACCIPITRIFORMES" "ACCIPITRIFORMES" "ACCIPITRIFORMES" ...  
## $ Family : chr "Accipitridae" "Accipitridae" "Accipitridae" "Accipitridae" ...  
## $ MVZDatabase : chr "Accipiter badius" "Accipiter cooperii" "Accipiter gentilis" "Accipiter nisus" ...  
## $ Species : chr "Accipiter badius" "Accipiter cooperii" "Accipiter gentilis" "Accipiter nisus" ...  
## $ Asymmetry : num 0.1378 0.0937 0.1114 0.0808 0.0749 ...  
## $ Ellipticity : num 0.344 0.272 0.319 0.239 0.254 ...  
## $ AvgLength : num 3.86 4.9 5.99 4.04 3.87 ...  
## $ NumberOfImages: int 1 27 7 13 15 1 191 1 7 2 ...  
## $ NumberOfEggs : int 2 103 18 61 57 1 391 2 17 4 ...
```

Subset the full data frame to Order Galliformes, chickens and other fowl.

```
eggGalli = egg[egg$Order == "GALLIFORMES",]
```

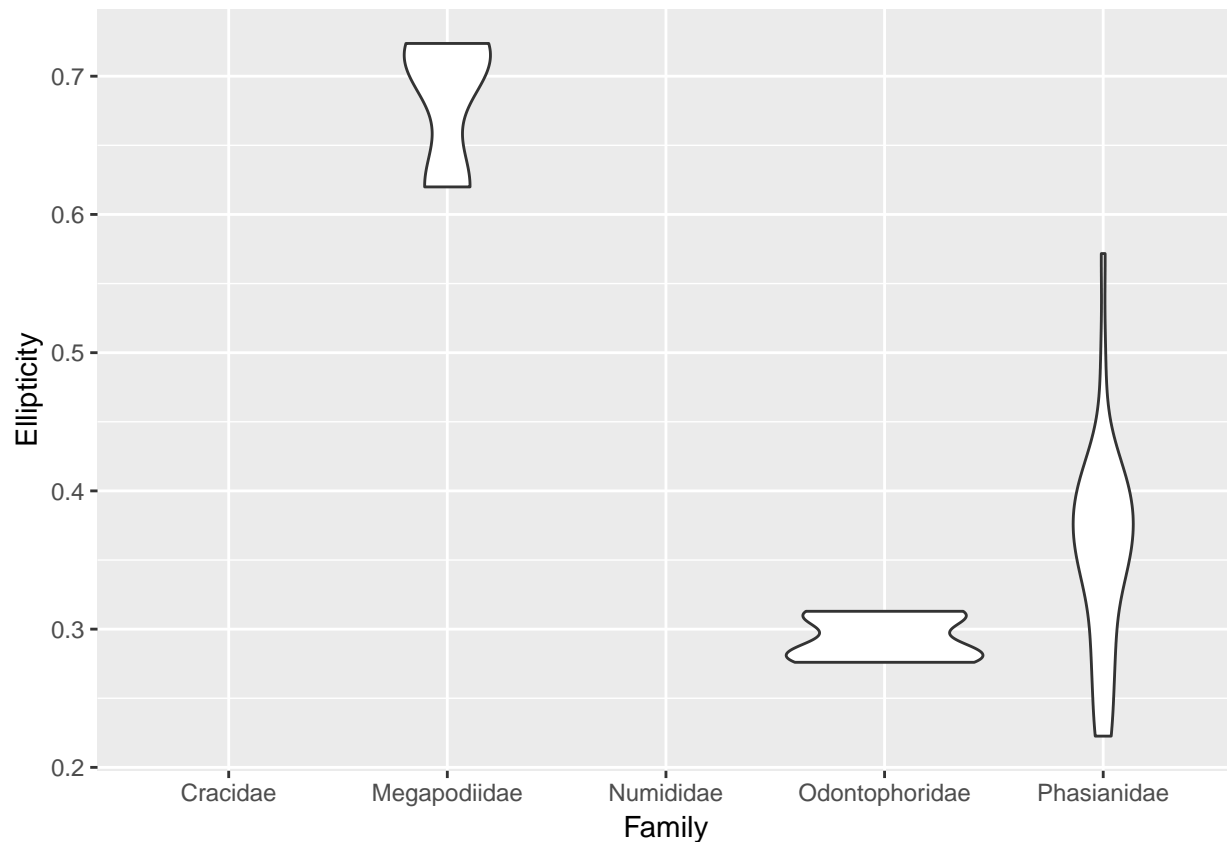
```
head(eggGalli)
```

```
##           Order           Family           MVZDatabase  
## 392 GALLIFORMES           Cracidae           Ortalis vetula  
## 393 GALLIFORMES Megapodiidae           Macrocephalon maleo  
## 394 GALLIFORMES Megapodiidae           Megapodius freycinet  
## 395 GALLIFORMES Megapodiidae           Megapodius pritchardii  
## 396 GALLIFORMES           Numididae           Numida meleagris  
## 397 GALLIFORMES Odontophoridae           Callipepla californica  
##           Species Asymmetry Ellipticity AvgLength NumberOfImages  
## 392           Ortalis vetula           0.1178           0.4590           6.0847           10  
## 393           Macrocephalon maleo           0.0365           0.7237           8.3400           1  
## 394           Megapodius freycinet           0.0551           0.6199           8.9621           1  
## 395           Megapodius pritchardii           0.0221           0.7072           7.9284           1  
## 396           Numida meleagris           0.2260           0.3197           5.2572           1  
## 397           Callipepla californica           0.2364           0.2850           3.0381           55
```

```
##      NumberOfEggs
## 392           31
## 393            1
## 394            2
## 395            3
## 396           10
## 397           700
```

## Violin plot

```
ggplot(data=eggGalli, mapping=aes(x=Family, y=Ellipticity)) +
  geom_violin()
```



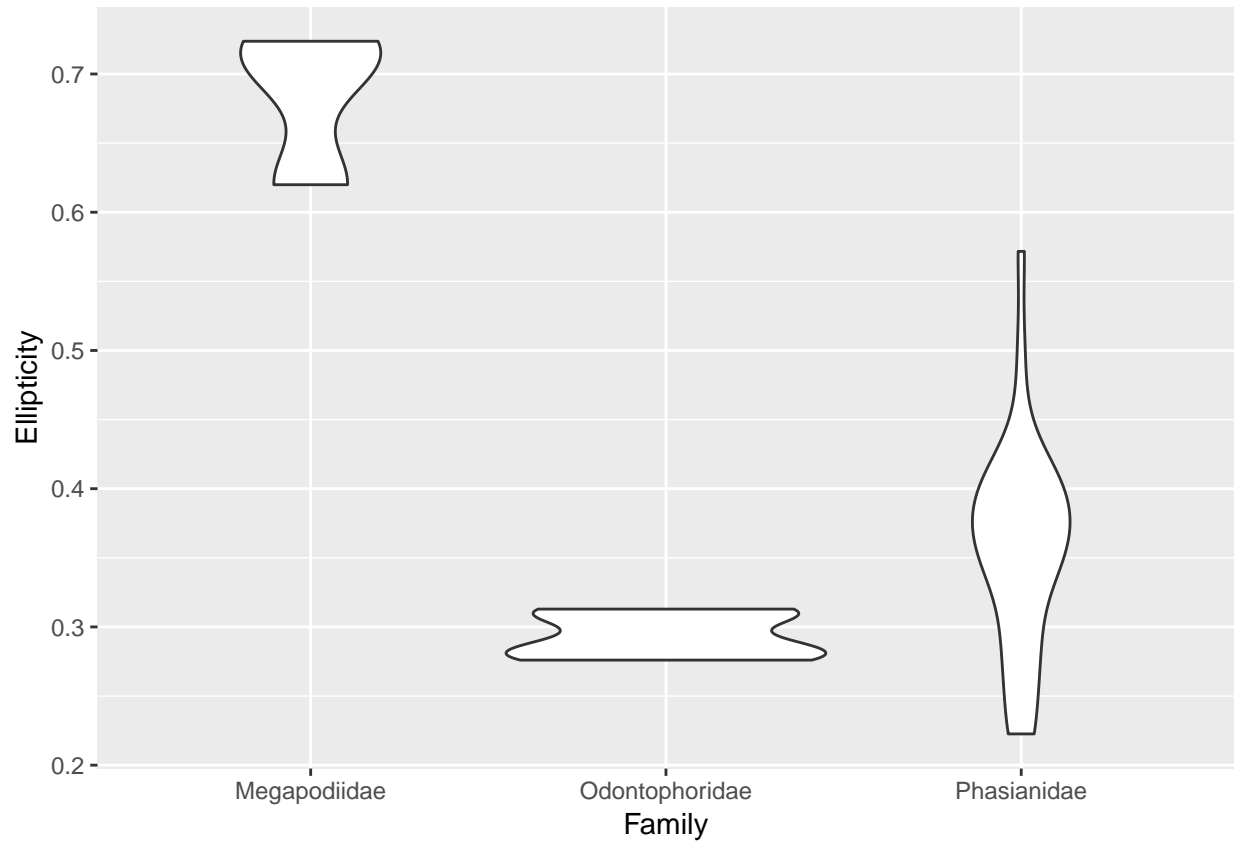
```
table(eggGalli$Family)
```

```
##
##      Cracidae  Megapodiidae  Numididae Odontophoridae  Phasianidae
##           1             3           1             7           36
```

Cracidae and Numididae don't show distributions because they each only have 1 observation.

We can subset the plot to not show them. Alternatively, we could add a `geom_point()` layer to show them as single points.

```
ggplot(data=eggGalli[eggGalli$Family != "Cracidae" &
  eggGalli$Family != "Numididae",],
  mapping=aes(x=Family, y=Ellipticity)) +
  geom_violin()
```



Our axis doesn't start at zero, which can be misleading.

```
range(egg$Ellipticity)
## [1] 0.0967 0.7237
```

Looks like the range of possible values is probably 0 to 1. Let's change our axis, and add the `expand` argument to get rid of the extra space beyond the axis limits.

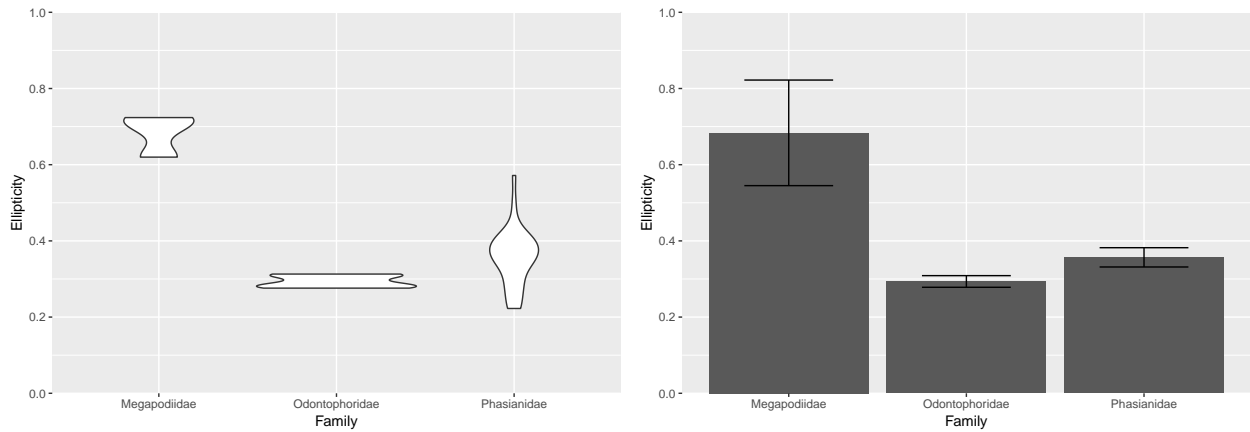
```
ggplot(data=eggGalli[eggGalli$Family != "Cracidae" &
  eggGalli$Family != "Numididae",],
  mapping=aes(x=Family, y=Ellipticity)) +
  geom_violin() +
  scale_y_continuous(breaks=seq(0,1,0.2),
    limits=c(0,1),
    expand=c(0,0))
```



Compare this to a bar plot of the same data:

```
ggplot(data=eggGalli[eggGalli$Family != "Cracidae" &
  eggGalli$Family != "Numididae",],
  mapping=aes(x=Family, y=Ellipticity)) +
  geom_violin() +
  scale_y_continuous(breaks=seq(0,1,0.2),
    limits=c(0,1),
    expand=c(0,0))

ggplot(data=eggGalli[eggGalli$Family != "Cracidae" &
  eggGalli$Family != "Numididae",],
  mapping=aes(x=Family, y=Ellipticity)) +
  geom_bar(stat="summary", fun.y="mean") +
  geom_errorbar(stat="summary", fun.data="mean_cl_normal",
    width=0.5) +
  scale_y_continuous(breaks=seq(0,1,0.2),
    limits=c(0,1),
    expand=c(0,0))
```



If we want, we can take the best of both and lay the confidence intervals on top of the violin plot:

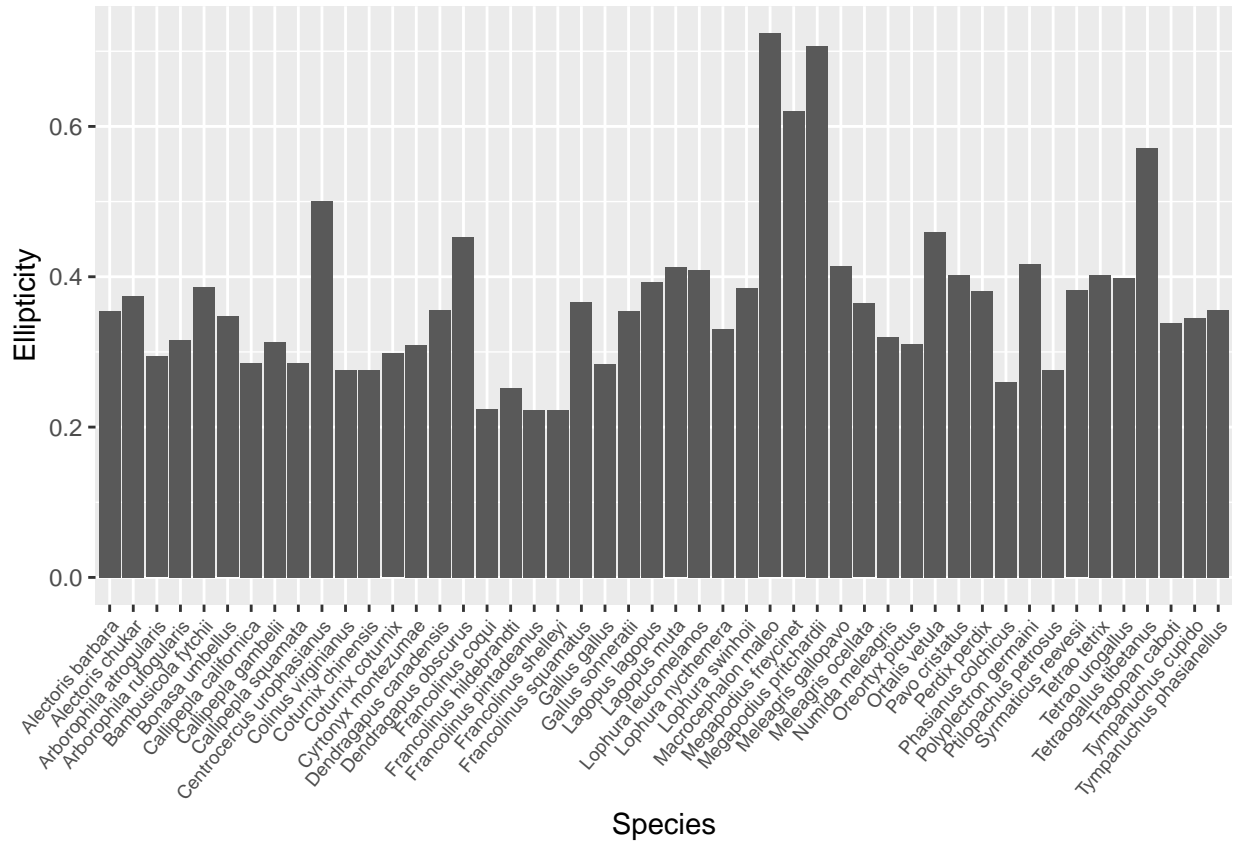
```
ggplot(data=eggGalli[eggGalli$Family != "Cracidae" &
  eggGalli$Family != "Numididae",],
  mapping=aes(x=Family, y=Ellipticity)) +
  geom_violin() +
  geom_errorbar(stat="summary", fun.data="mean_cl_normal",
    width=0.1) +
  scale_y_continuous(breaks=seq(0,1,0.2),
    limits=c(0,1),
    expand=c(0,0))
```



## Facets

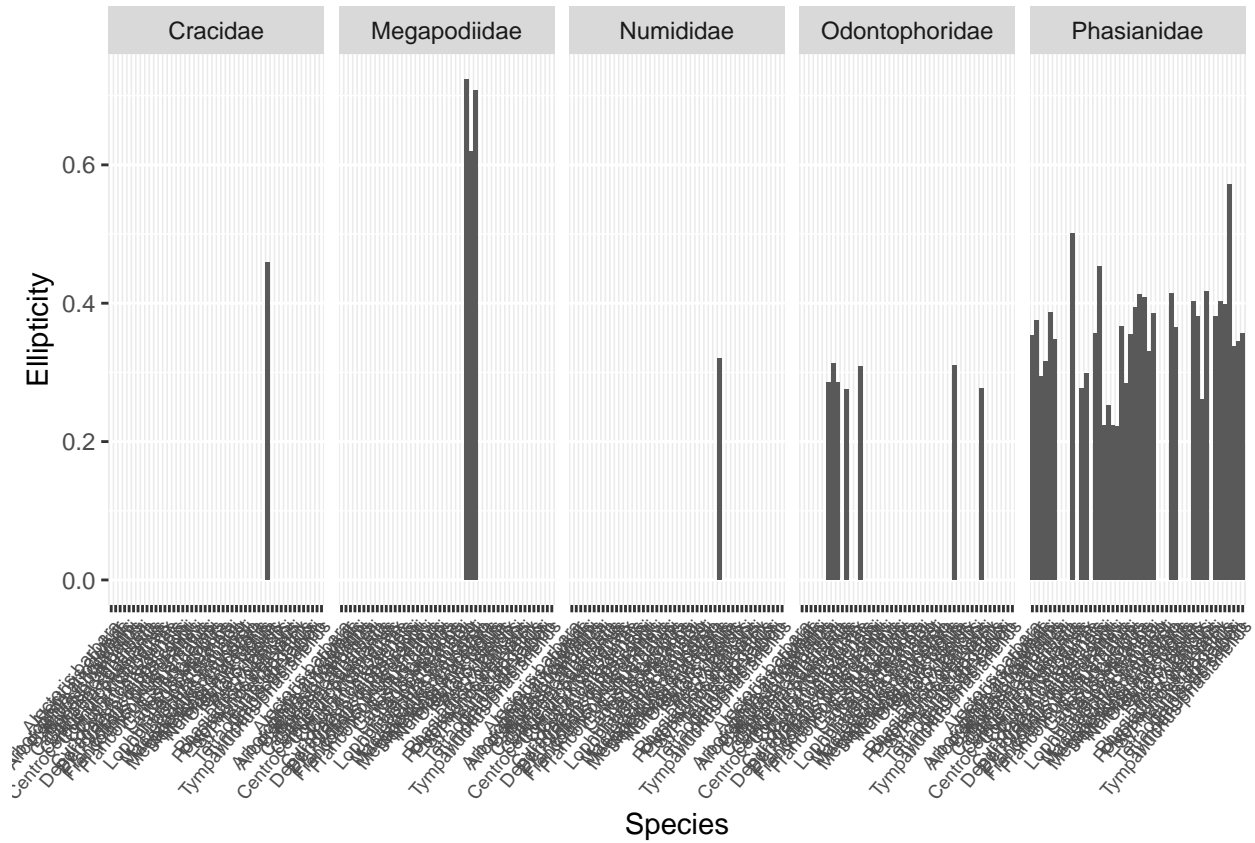
Plot all ellipticity values by species.

```
ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +  
  geom_bar(stat="identity") +  
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))
```



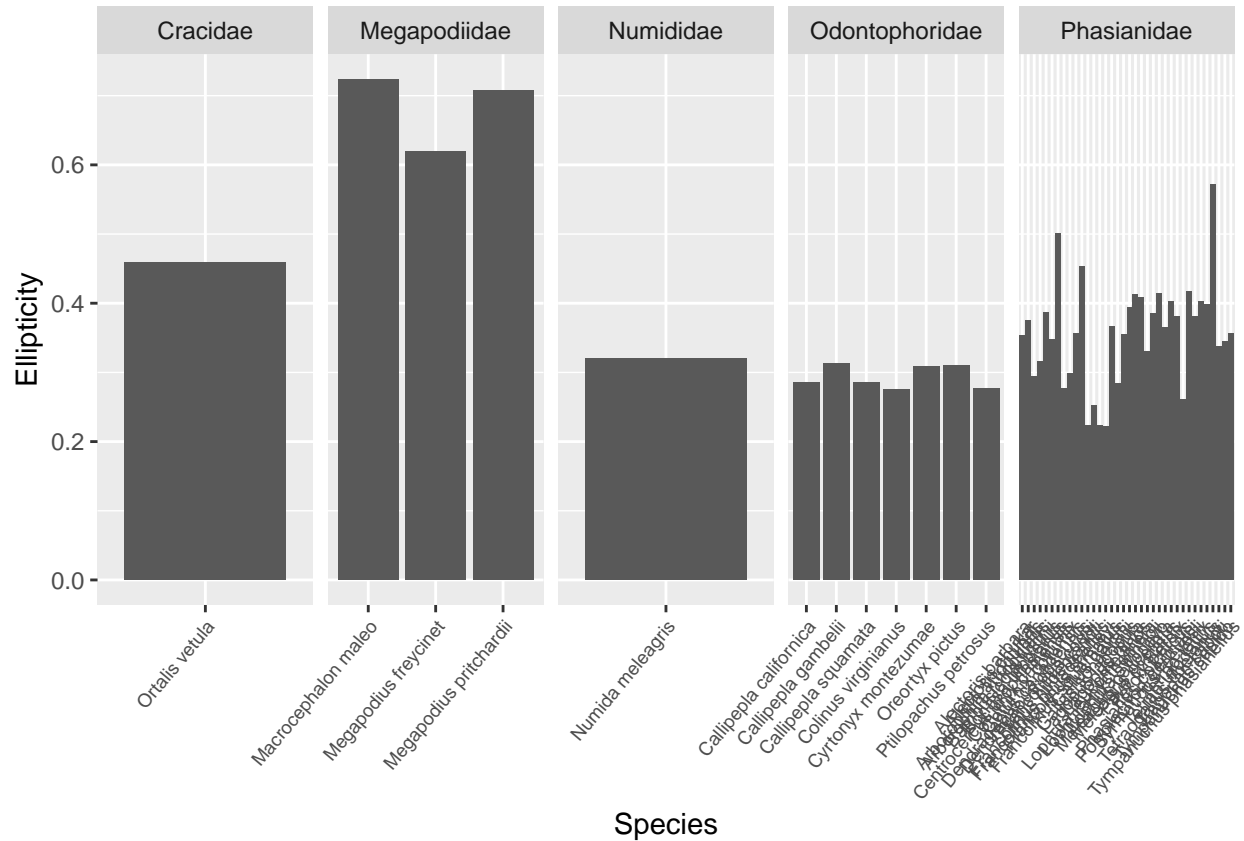
Add a facet to group them by family.

```
ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +  
  facet_grid(cols=vars(Family)) +  
  geom_bar(stat="identity") +  
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))
```



... But all of our values appear under each facet. Not what we want. So we add `scales="free"` to allow scales to vary across rows and columns.

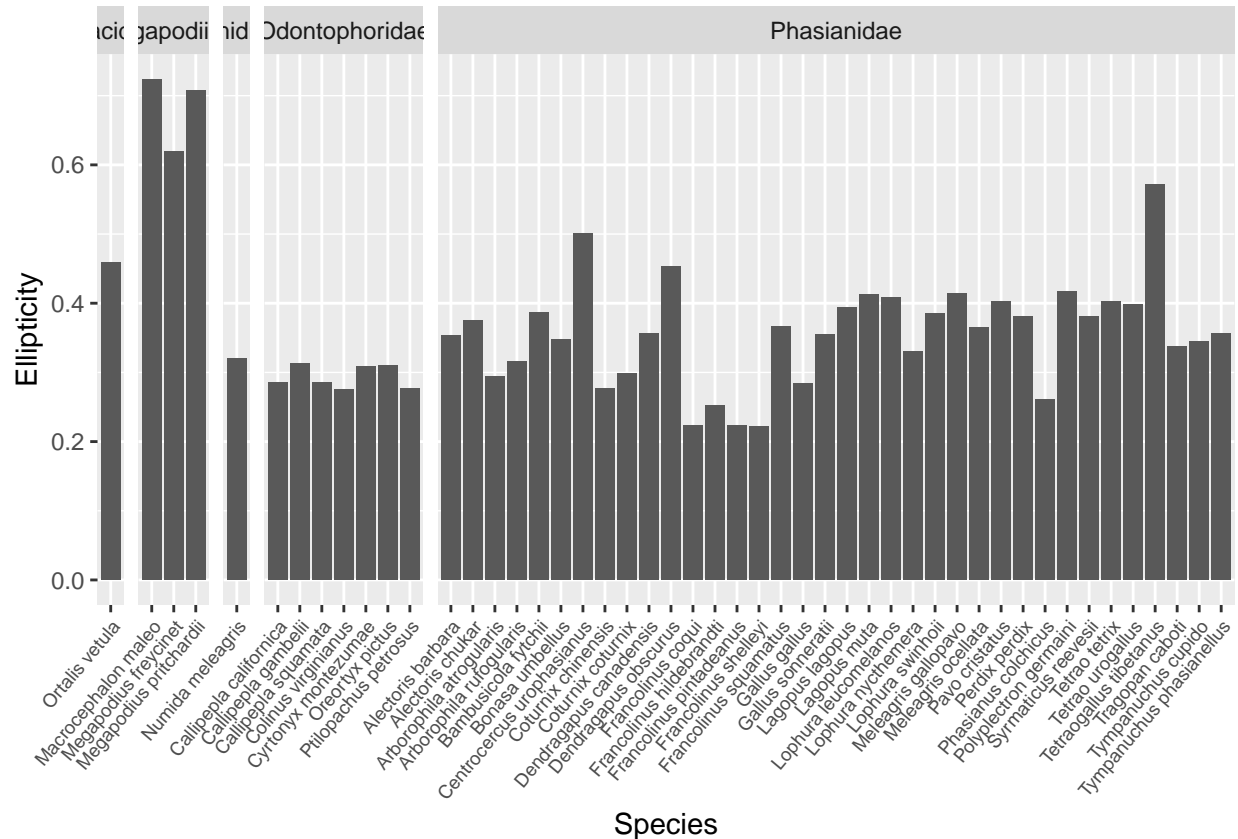
```
ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free") +
  geom_bar(stat="identity") +
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))
```



Still not quite right, as some bars are now wider than others and may seem like they're communicating some other information. So we add `space="free"` to allow panel sizes to vary.

```
ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free", space="free") +
  geom_bar(stat="identity") +
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))
```





And we've now made a new problem in that our facet labels don't fit anymore. So let's use the `recode()` function from `dplyr` to change those values.

```
unique(eggGalli$Family)

## [1] "Cracidae"          "Megapodiidae"     "Numididae"       "Odontophoridae"
## [5] "Phasianidae"

library(dplyr)

##
## Attaching package: 'dplyr'

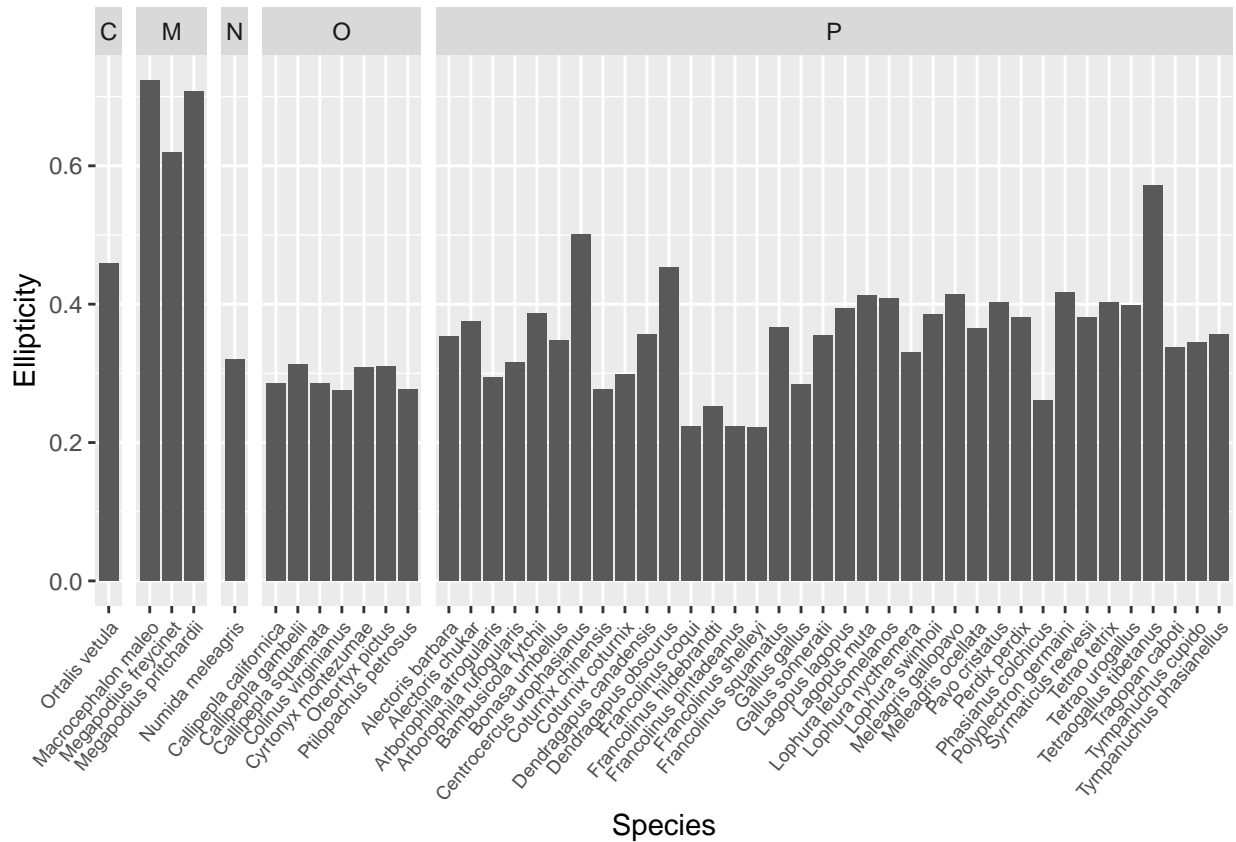
## The following objects are masked from 'package:stats':
##
##   filter, lag

## The following objects are masked from 'package:base':
##
##   intersect, setdiff, setequal, union

eggGalli$Family = recode(eggGalli$Family,
  Cracidae = "C",
  Megapodiidae = "M",
  Numididae = "N",
  Odontophoridae = "O",
  Phasianidae = "P")

ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free", space="free") +
```

```
geom_bar(stat="identity") +
theme(axis.text.x=element_text(angle=50, hjust=1, size=7))
```



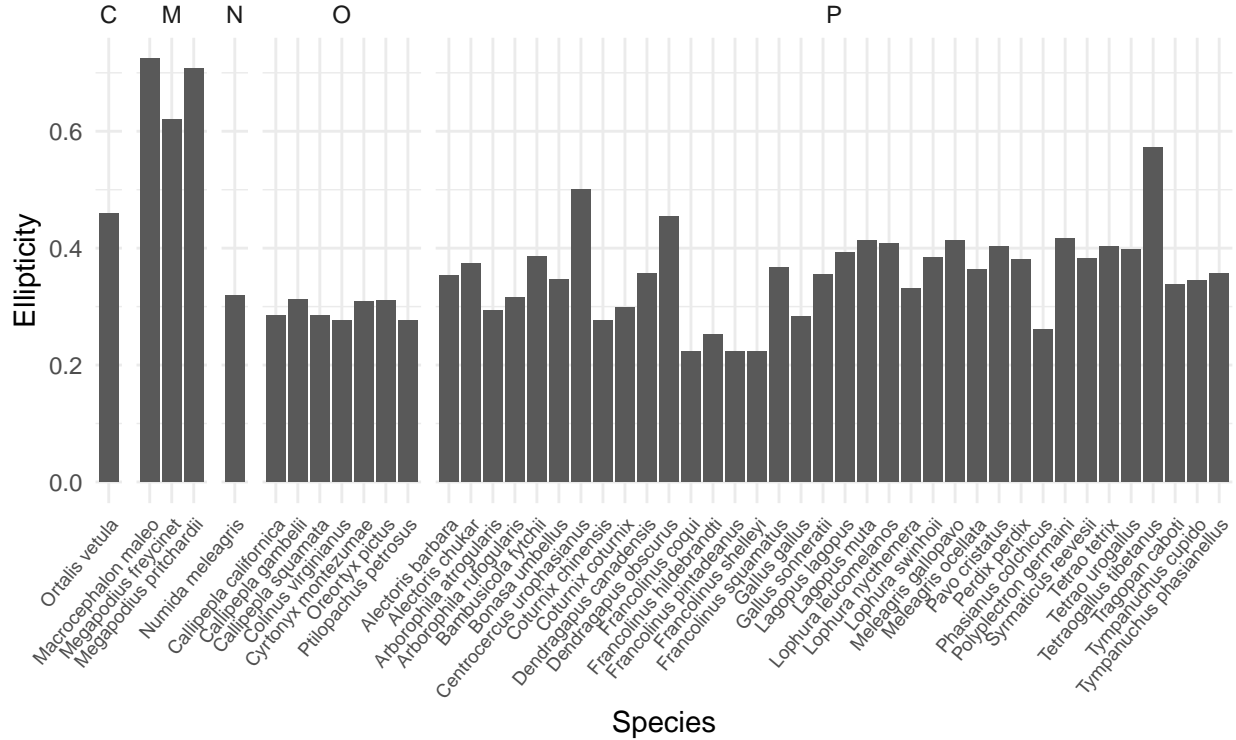
## Themes

`theme_gray()` is the default

`theme_minimal()`

```
ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free", space="free") +
  geom_bar(stat="identity") +
  labs(title="Egg ellipticity by family in Galliformes spp.",
        caption="Data from Stoddard et al. (2017)") +
  theme_minimal() +
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))
```

## Egg ellipticity by family in Galliformes spp.



Data from Stoddard et al. (2017)

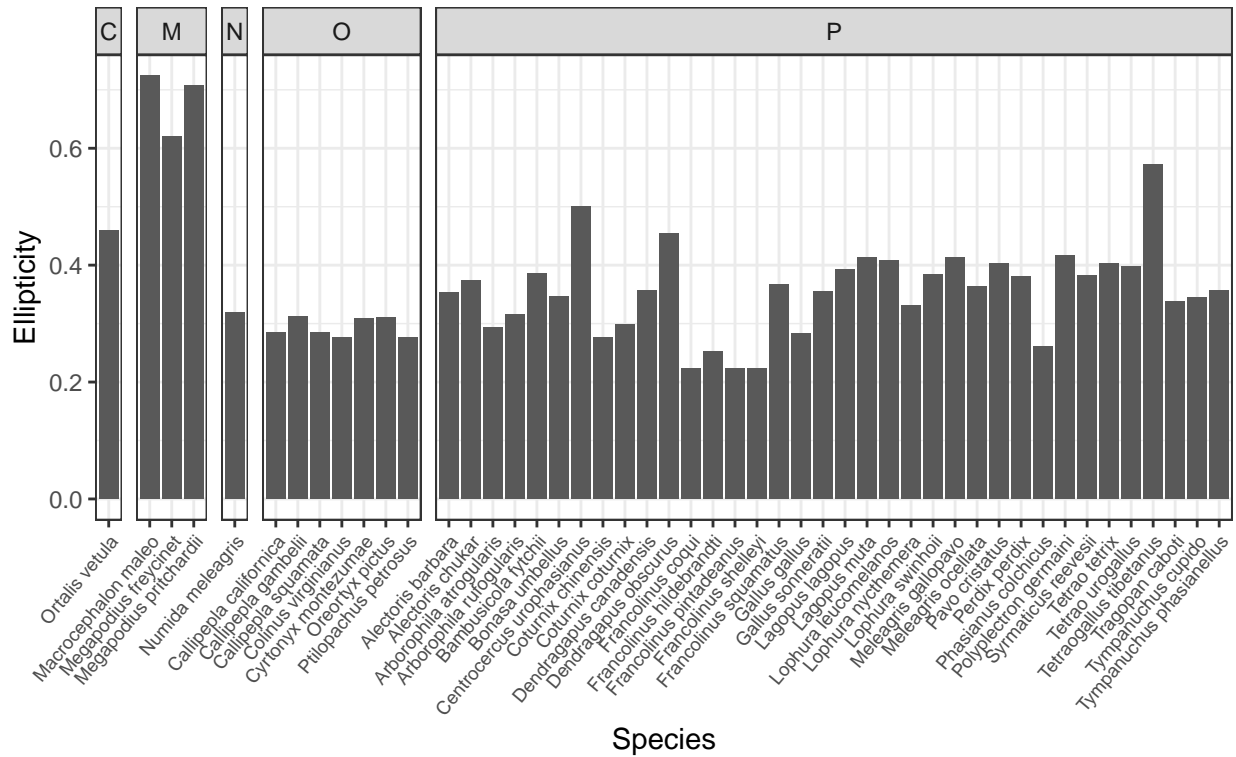
```

theme_bw()

ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free", space="free") +
  geom_bar(stat="identity") +
  labs(title="Egg ellipticity by family in Galliformes spp.",
        caption="Data from Stoddard et al. (2017)") +
  theme_bw() +
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))

```

### Egg ellipticity by family in Galliformes spp.



Data from Stoddard et al. (2017)

```

theme_dark()

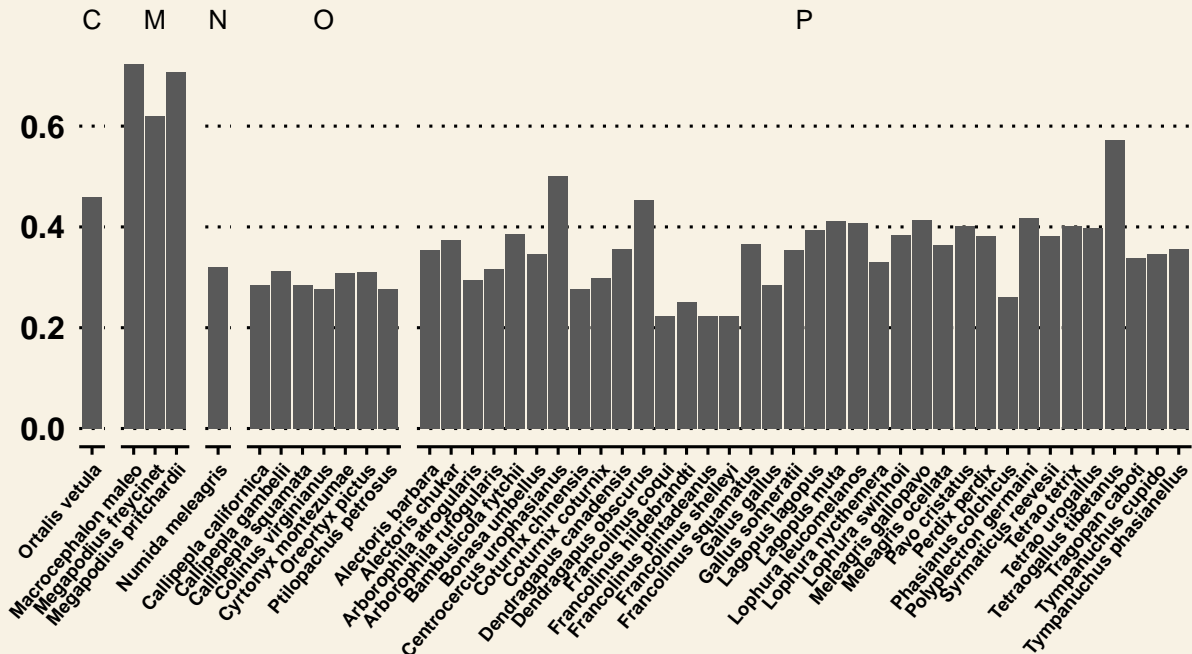
ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free", space="free") +
  geom_bar(stat="identity") +
  labs(title="Egg ellipticity by family in Galliformes spp.",
        caption="Data from Stoddard et al. (2017)") +
  theme_dark() +
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))

```





# Egg ellipticity by family

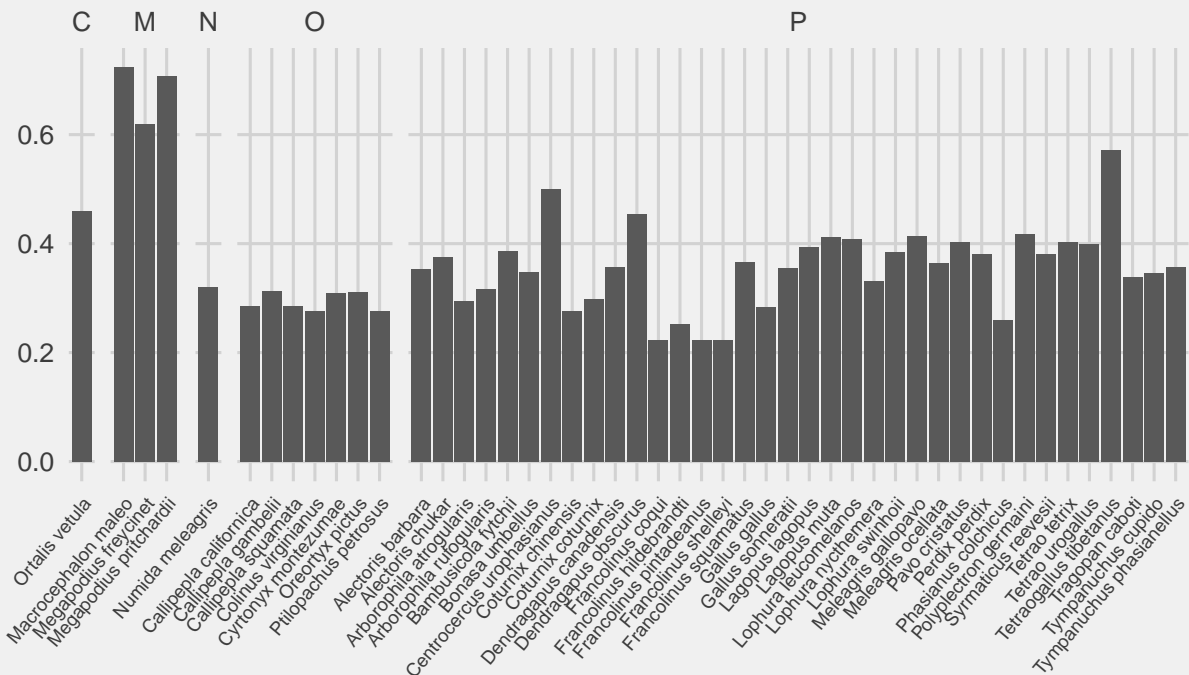


Data from Stoddard et al. (2017)

```
ggthemes::theme_fivethirtyeight()

ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free", space="free") +
  geom_bar(stat="identity") +
  labs(title="Egg ellipticity by family in Galliformes spp.",
       caption="Data from Stoddard et al. (2017)") +
  theme_fivethirtyeight() +
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))
```

## Egg ellipticity by family in Galliformes spp.

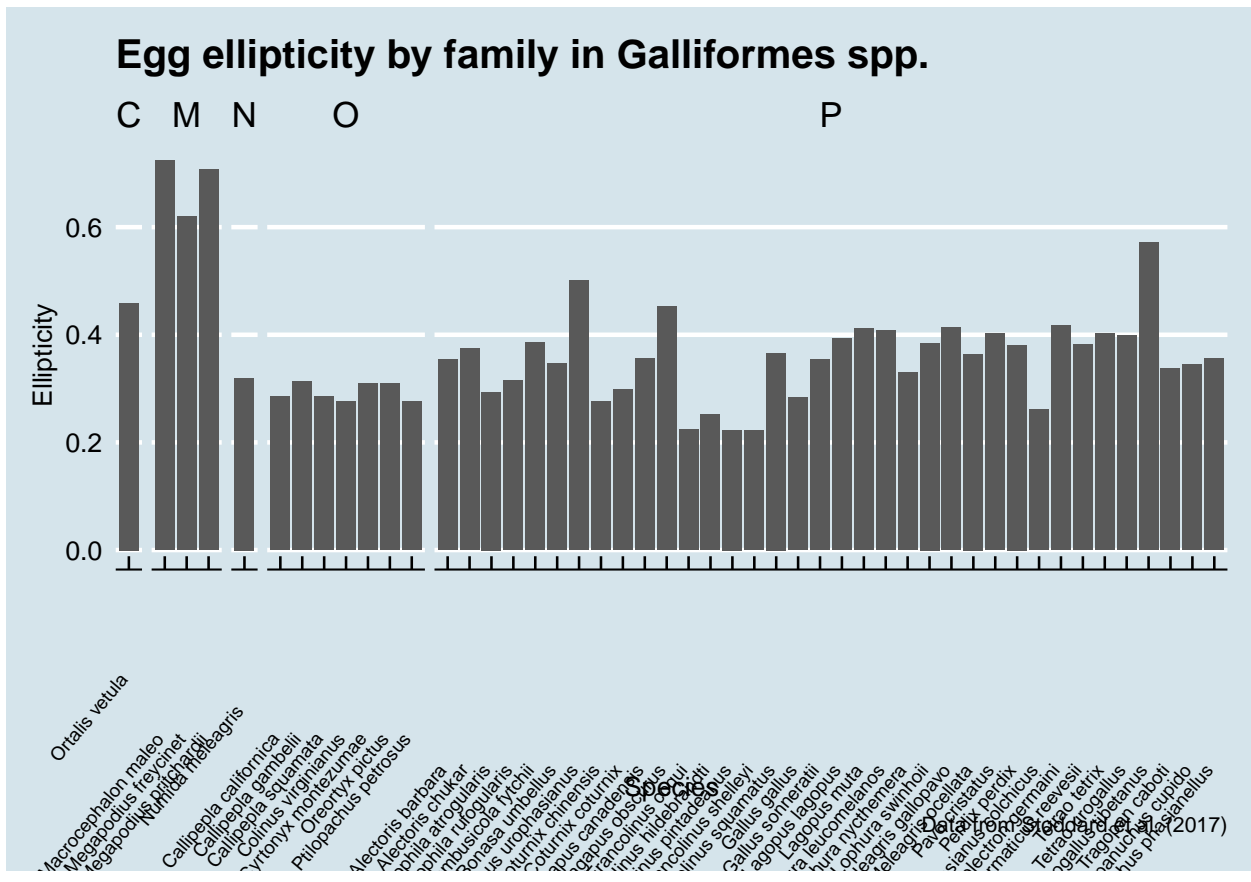


Data from Stoddard et al. (2017)

```
ggthemes::theme_economist()

ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free", space="free") +
  geom_bar(stat="identity") +
  labs(title="Egg ellipticity by family in Galliformes spp.",
       caption="Data from Stoddard et al. (2017)") +
  theme_economist() +
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))
```





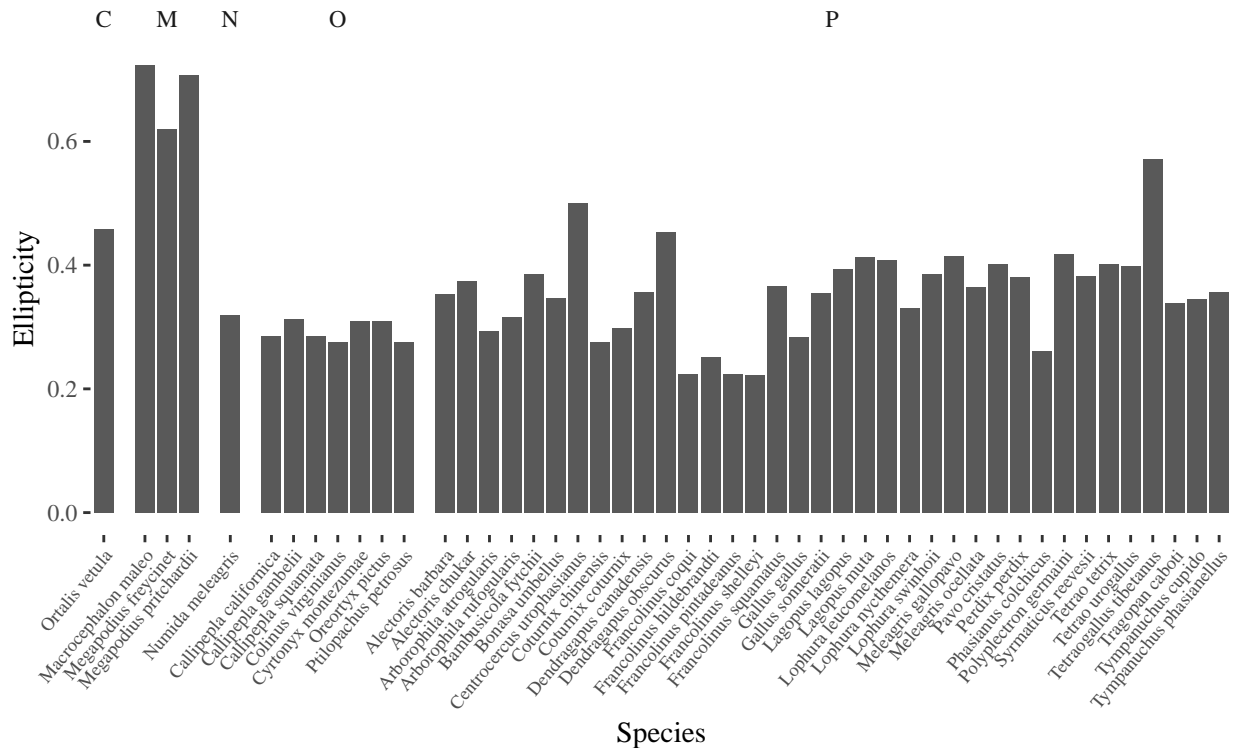
```

ggthemes::theme_tufte()

ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free", space="free") +
  geom_bar(stat="identity") +
  labs(title="Egg ellipticity by family in Galliformes spp.",
        caption="Data from Stoddard et al. (2017)") +
  theme_tufte() +
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7))

```

## Egg ellipticity by family in Galliformes spp.



Data from Stoddard et al. (2017)

Or, for fun, xkcd... (like the webcomic)

```
install.packages("xkcd")
install.packages("extrafont")

library(xkcd)

## Loading required package: extrafont

## Registering fonts with R

download.file("http://simonsoftware.se/other/xkcd.ttf",
             dest="./resources/xkcd.ttf", mode="wb")
font_import(paths="./resources/", pattern="[X/x]kcd", prompt=FALSE)
loadfonts(device="win") # For Mac: loadfonts()

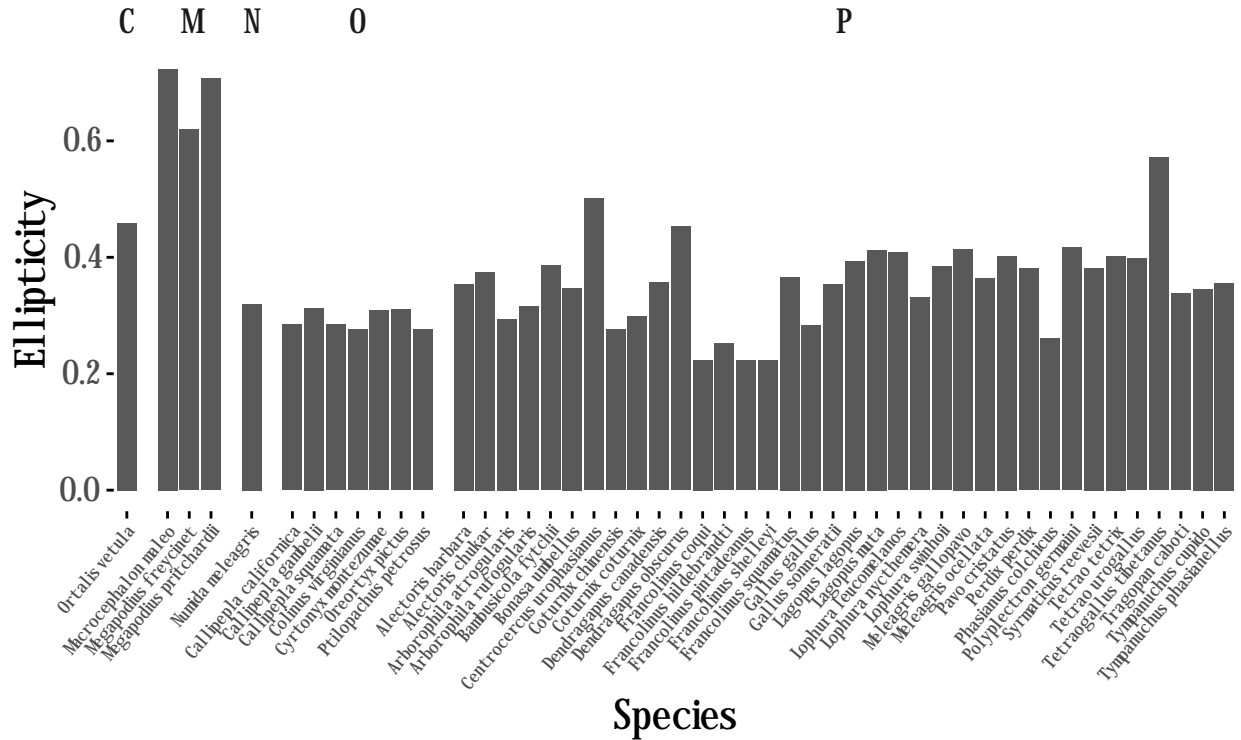
# Also have to put the font in your system fonts directory

ggplot(data=eggGalli, mapping=aes(x=Species, y=Ellipticity)) +
  facet_grid(cols=vars(Family), scales="free", space="free") +
  geom_bar(stat="identity") +
  labs(title="Egg ellipticity by family in Galliformes spp.",
       caption="Data from Stoddard et al. (2017)") +
  theme_xkcd() +
  theme(axis.text.x=element_text(angle=50, hjust=1, size=7),
        text=element_text(family="xkcd"))

## Warning in grid.Call(C_textBounds, as.graphicsAnnot(x$label), x$x, x$y, :
## font width unknown for character 0x28
```



# Egg ellipticity by family in Galliformes spp.



Data from Stoddard et al. (2017)

There are additional functions to make “fuzzy” rectangles and lines, but they’re very particular about the input so you’d have to reformat your data:

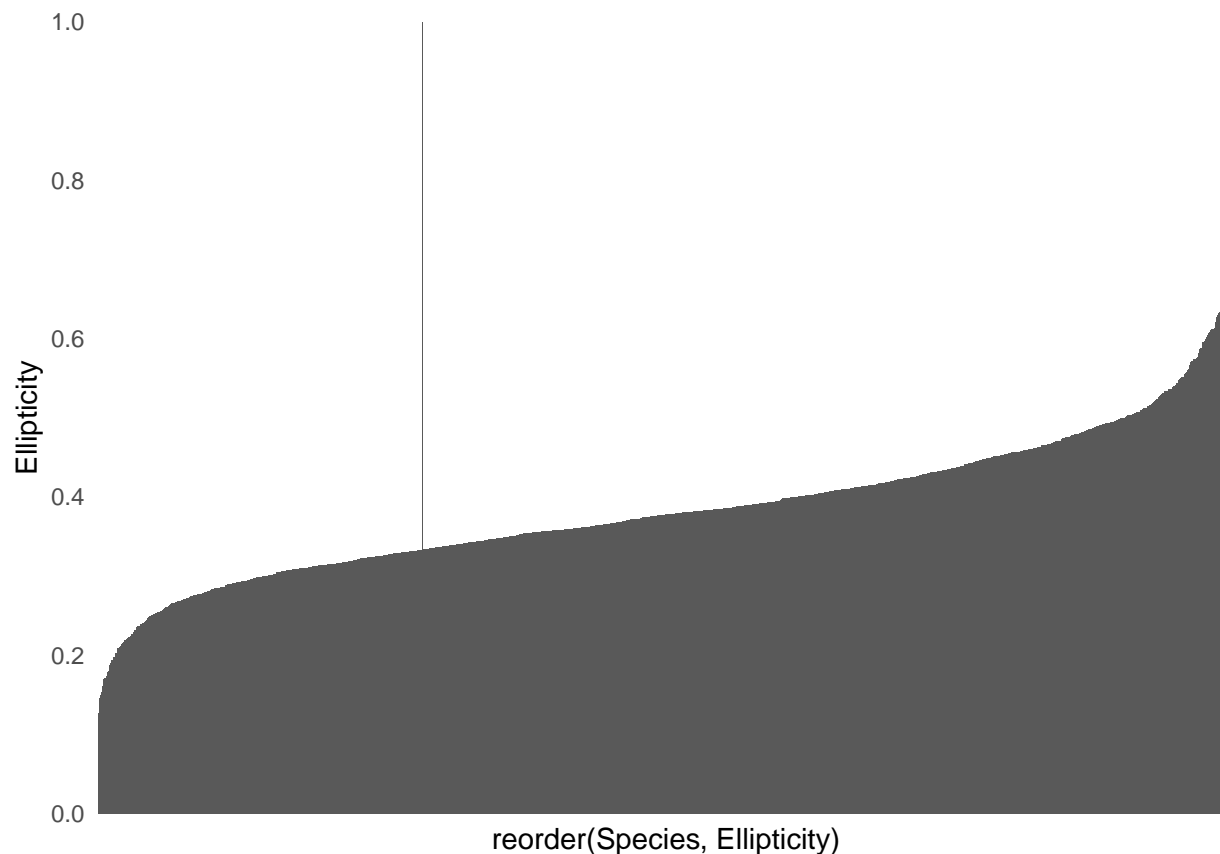
```
?xkcdirect
?xkcdline
```

## Color scales

Arrange all species in the original data frame by their ellipticity values (and remove the grid and tick marks):

```
ggplot(data=egg, mapping=aes(x=reorder(Species, Ellipticity),
                               y=Ellipticity)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,1,0.2),
                    limits=c(0,1),
                    expand=c(0,0)) +
  theme_minimal() +
  theme(axis.text.x=element_blank(),
        panel.grid.major=element_blank(),
        panel.grid.minor=element_blank())

## Warning: Removed 2 rows containing missing values (geom_bar).
```



We're seeing a warning about missing values.

```
egg[is.na(egg$Species),]
```

```
##      Order      Family      MVZDatabase Species Asymmetry
## 377 EXTINCT Aepyornithidae      Aepyornis sp. <NA> 0.0044
## 378 EXTINCT      Columbidae Ectopistes migratorius <NA> 0.0872
## 379 EXTINCT Dinornithidae      Dinornis sp. <NA> 0.0050
## 380 EXTINCT      Rallidae      Porzana palmeri <NA> 0.0523
##      Ellipticity AvgLength NumberOfImages NumberOfEggs
## 377      0.4499    23.8700             2             2
## 378      0.3391     3.9290             5             6
## 379      0.3318    14.4235             1             1
## 380      0.4494     2.9990             1             1
```

There are some extinct species without a `Species` value. To get rid of the warning, we can fix it for now by using the `MVZDatabase` values, which are good enough:

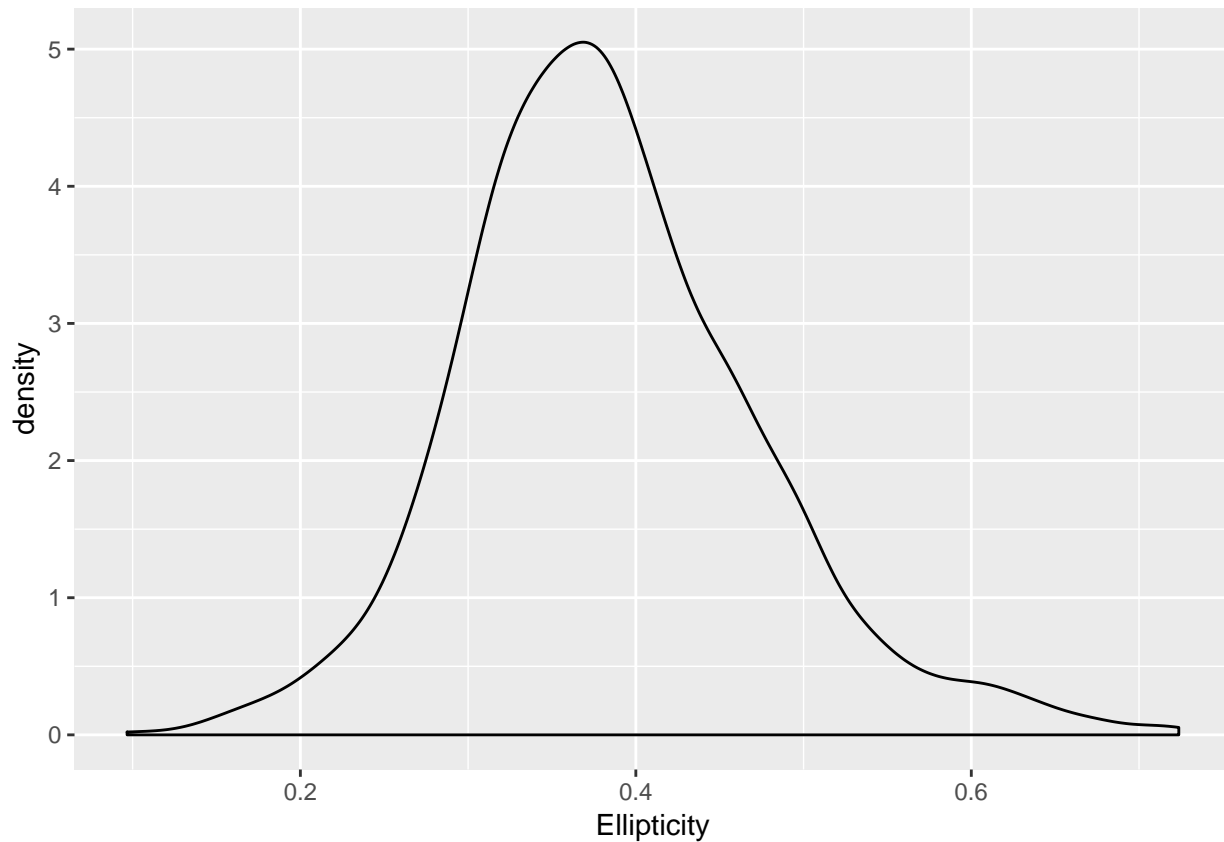
```
egg$Species[is.na(egg$Species)] = egg$MVZDatabase[is.na(egg$Species)]
```

```
egg[is.na(egg$Species),]
```

```
## [1] Order      Family      MVZDatabase  Species
## [5] Asymmetry    Ellipticity AvgLength    NumberOfImages
## [9] NumberOfEggs
## <0 rows> (or 0-length row.names)
```

We also saw a beautiful sigmoidal curve for the ellipticity values when we lined them all up. Why?

```
ggplot(data=egg, mapping=aes(x=Ellipticity)) +
  geom_density()
```



Because the distribution of values is really close to a “normal” Gaussian curve.

Now let’s subset the data for Order Cuculiformes, the cuckoos.

```
eggCuckoo = egg[egg$Order == "CUCULIFORMES",]
eggCuckoo$Species
```

```
## [1] "Centropus bengalensis"      "Centropus senegalensis"
## [3] "Centropus superciliosus"    "Centropus viridis"
## [5] "Coccyzus americanus"       "Coccyzus erythrophthalmus"
## [7] "Coccyzus minor"           "Crotophaga ani"
## [9] "Crotophaga sulcirostris"    "Cuculus pallidus"
## [11] "Geococcyx californianus"    "Geococcyx velox"
## [13] "Guira guira"                "Phaenicophaeus superciliosus"
```

We also want to take these Species values and split them to make a Genus variable too (since we have Order, Family, and Species, but not Genus).

We do this by using `strsplit()` and a fancy `apply()`-family function:

```
strsplit(eggCuckoo$Species, " ")
## [[1]]
## [1] "Centropus"  "bengalensis"
##
## [[2]]
## [1] "Centropus"  "senegalensis"
```

```

##
## [[3]]
## [1] "Centropus"      "superciliosus"
##
## [[4]]
## [1] "Centropus" "viridis"
##
## [[5]]
## [1] "Coccyzus"      "americanus"
##
## [[6]]
## [1] "Coccyzus"      "erythroptthalmus"
##
## [[7]]
## [1] "Coccyzus" "minor"
##
## [[8]]
## [1] "Crotophaga" "ani"
##
## [[9]]
## [1] "Crotophaga" "sulcirostris"
##
## [[10]]
## [1] "Cuculus" "pallidus"
##
## [[11]]
## [1] "Geococcyx"      "californianus"
##
## [[12]]
## [1] "Geococcyx" "velox"
##
## [[13]]
## [1] "Guira" "guira"
##
## [[14]]
## [1] "Phaenicophaeus" "superciliosus"

eggCuckoo$Genus = sapply(strsplit(eggCuckoo$Species, " "), "[", 1)
unique(eggCuckoo$Genus)

## [1] "Centropus"      "Coccyzus"      "Crotophaga"      "Cuculus"
## [5] "Geococcyx"      "Guira"          "Phaenicophaeus"

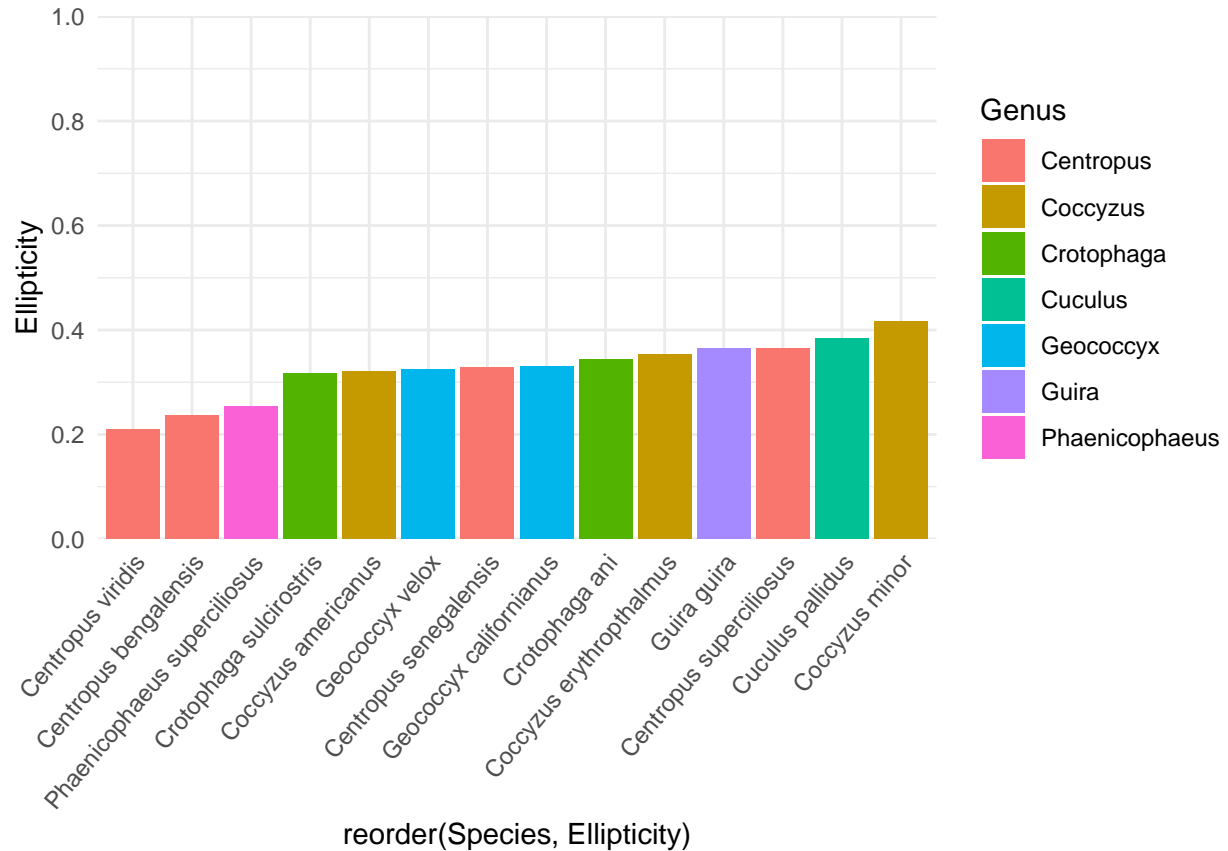
```

Now let's plot the cuckoo species, ordered by ellipticity values, with their fill colors according to genus.

```

ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                     y=Ellipticity,
                                     fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,1,0.2),
                    limits=c(0,1),
                    expand=c(0,0)) +
  theme_minimal() +
  theme(axis.text.x=element_text(angle=50, hjust=1))

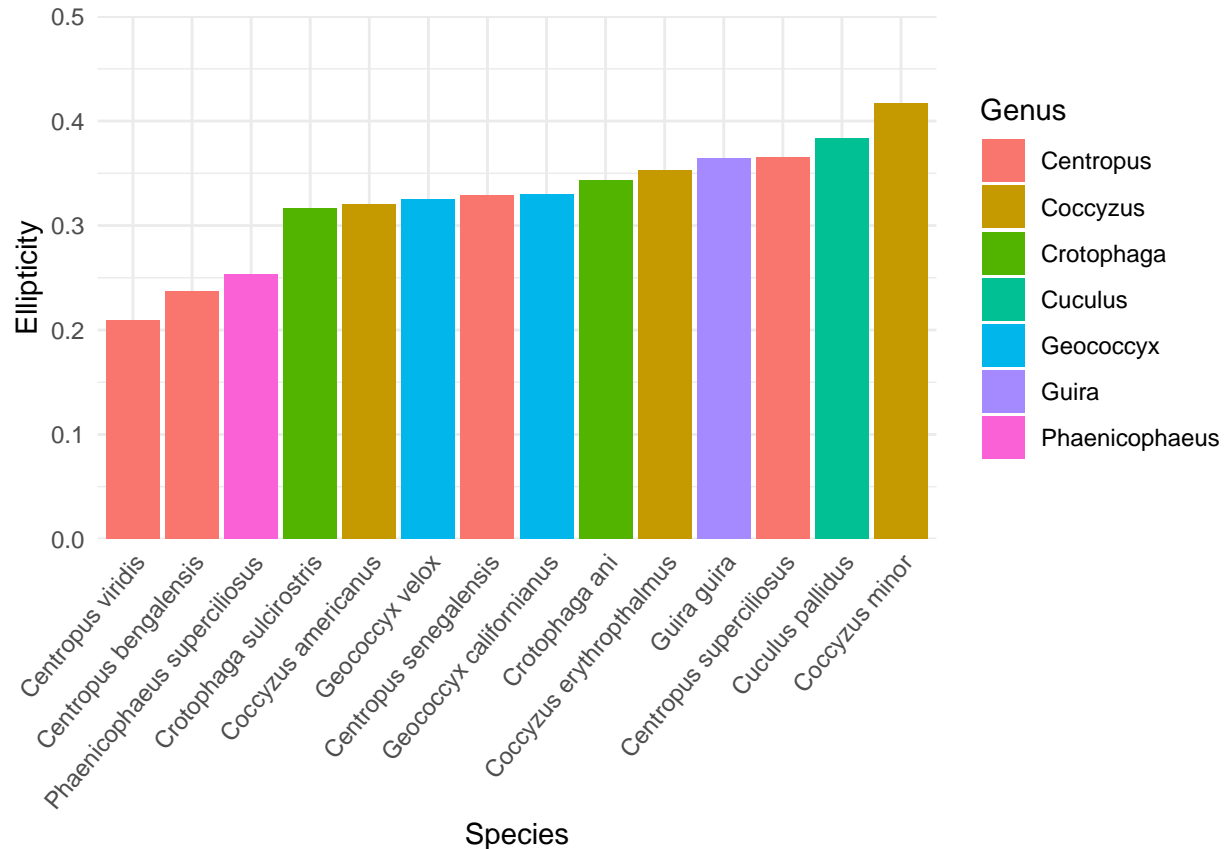
```



Let's readjust the Y axis and fix the X axis label.

```
ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                         y=Ellipticity,
                                         fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,0.5,0.1),
                    limits=c(0,0.5),
                    expand=c(0,0)) +
  labs(x="Species") +
  theme_minimal() +
  theme(axis.text.x=element_text(angle=50, hjust=1))
```





Now we want to customize the colors instead of using the default R palette.

```
length(unique(eggCuckoo$Genus))
```

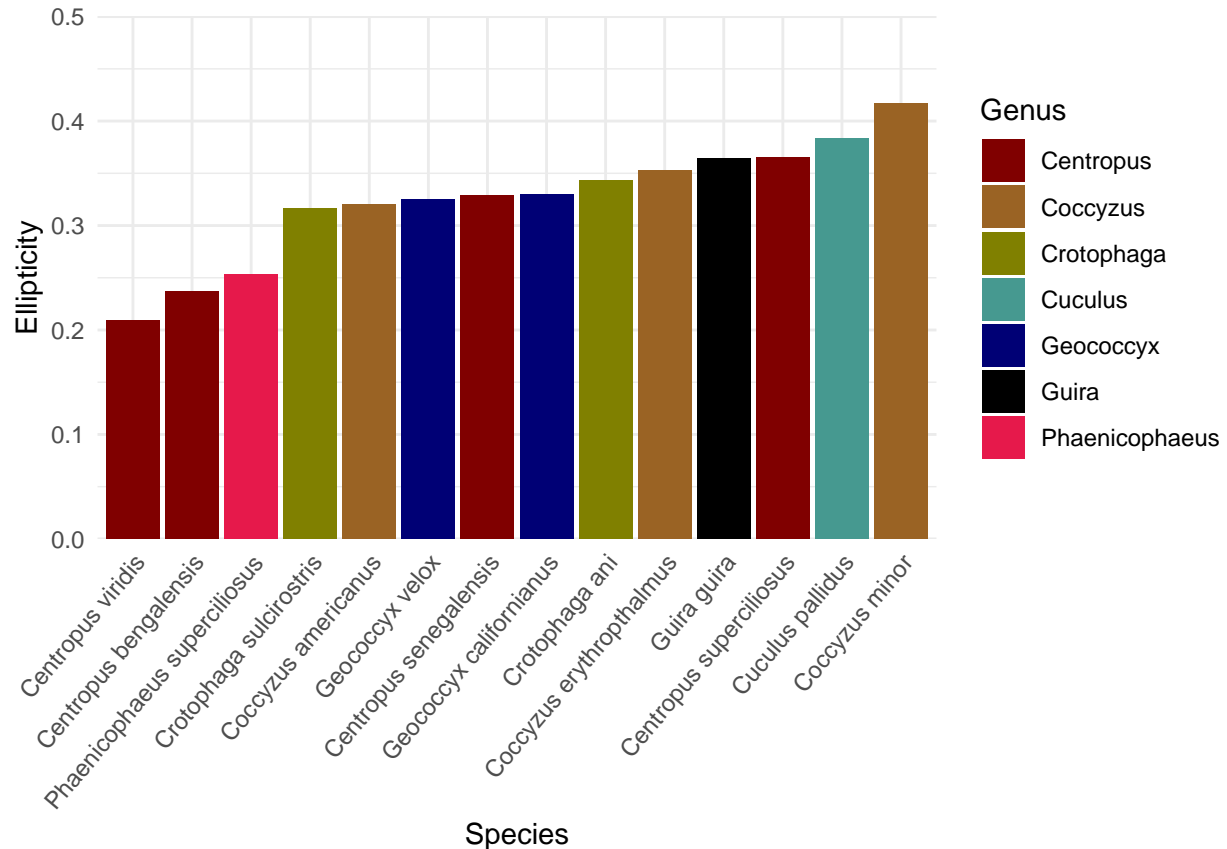
```
## [1] 7
```

We will need 7 different color values, and those values need to be for discrete or categorical data, not continuous (Ellipticity, for instance, would need a continuous scale).

## Manual

The first option is to assign color values manually, either by name or by hexadecimal value.

```
ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                     y=Ellipticity,
                                     fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,0.5,0.1),
                    limits=c(0,0.5),
                    expand=c(0,0)) +
  scale_fill_manual(values=c("#800000", "#9A6324", "#808000", "#469990",
                             "#000075", "#000000", "#e6194B")) +
  labs(x="Species") +
  theme_minimal() +
  theme(axis.text.x=element_text(angle=50, hjust=1))
```



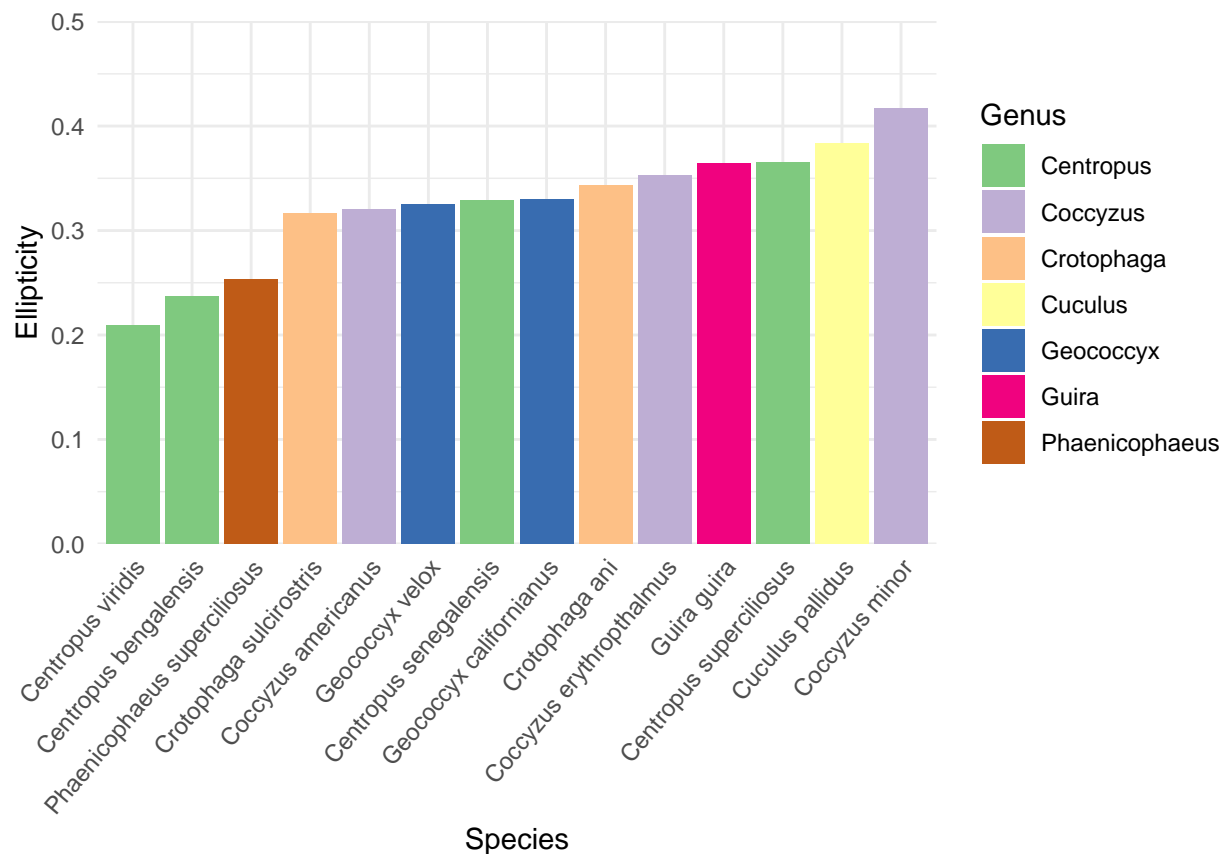
Source: Sasha Trubetskoy

### ColorBrewer

Or we can use the `brewer` scales that are included in `ggplot2` and can be viewed online at: <http://colorbrewer2.org/>

Here, we just specify the type of scale (qualitative) and the palette to be used (here, "Accent"). Other choices are shown in `?scale_fill_brewer`.

```
ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                     y=Ellipticity,
                                     fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,0.5,0.1),
                    limits=c(0,0.5),
                    expand=c(0,0)) +
  scale_fill_brewer(type="qual", palette="Accent") +
  labs(x="Species") +
  theme_minimal() +
  theme(axis.text.x=element_text(angle=50, hjust=1))
```



Source: ColorBrewer 2.0

### Color-blind friendly

But none of the palettes we've used so far are color-blind friendly, and may not be discernable in print or grayscale (after being photocopied), either. There are better palettes out there that are.

#### dichromat

```
install.packages("dichromat")
```

```
library(dichromat)
```

```
## Warning: package 'dichromat' was built under R version 3.5.2
```

The `dichromat()` function will take a list of colors (again, named or hexadecimal values) and convert them into a comparable color-blind friendly palette for a specific type of color-blindness (here: protanopia, the kind I have). When in doubt, you can use `deuteranopia`, the most common form of dichromacy.

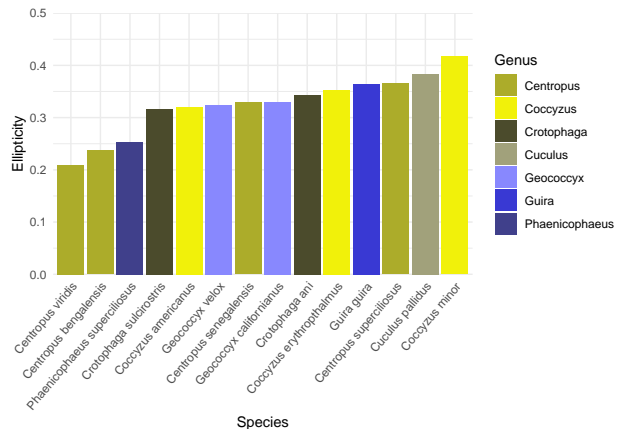
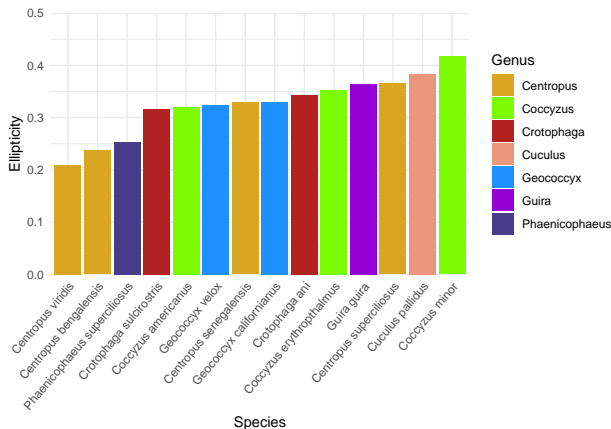
```
dichromat(c("goldenrod", "lawngreen", "firebrick",
            "darksalmon", "dodgerblue", "darkviolet",
            "darkslateblue"),
          type="protan")
```

```
## [1] "#ACAC2A" "#F1F10A" "#4B4B2C" "#A1A17B" "#8888FE" "#3939D3" "#40408B"
```

Compare the original colors (on the left) with the ones substituted by `dichromat()` (on the right):

```
ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                         y=Ellipticity,
                                         fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,0.5,0.1),
                    limits=c(0,0.5),
                    expand=c(0,0)) +
  scale_fill_manual(values=c("goldenrod","lawngreen","firebrick",
                             "darksalmon","dodgerblue","darkviolet",
                             "darkslateblue")) +
  labs(x="Species") +
  theme_minimal() +
  theme(axis.text.x=element_text(angle=50, hjust=1))
```

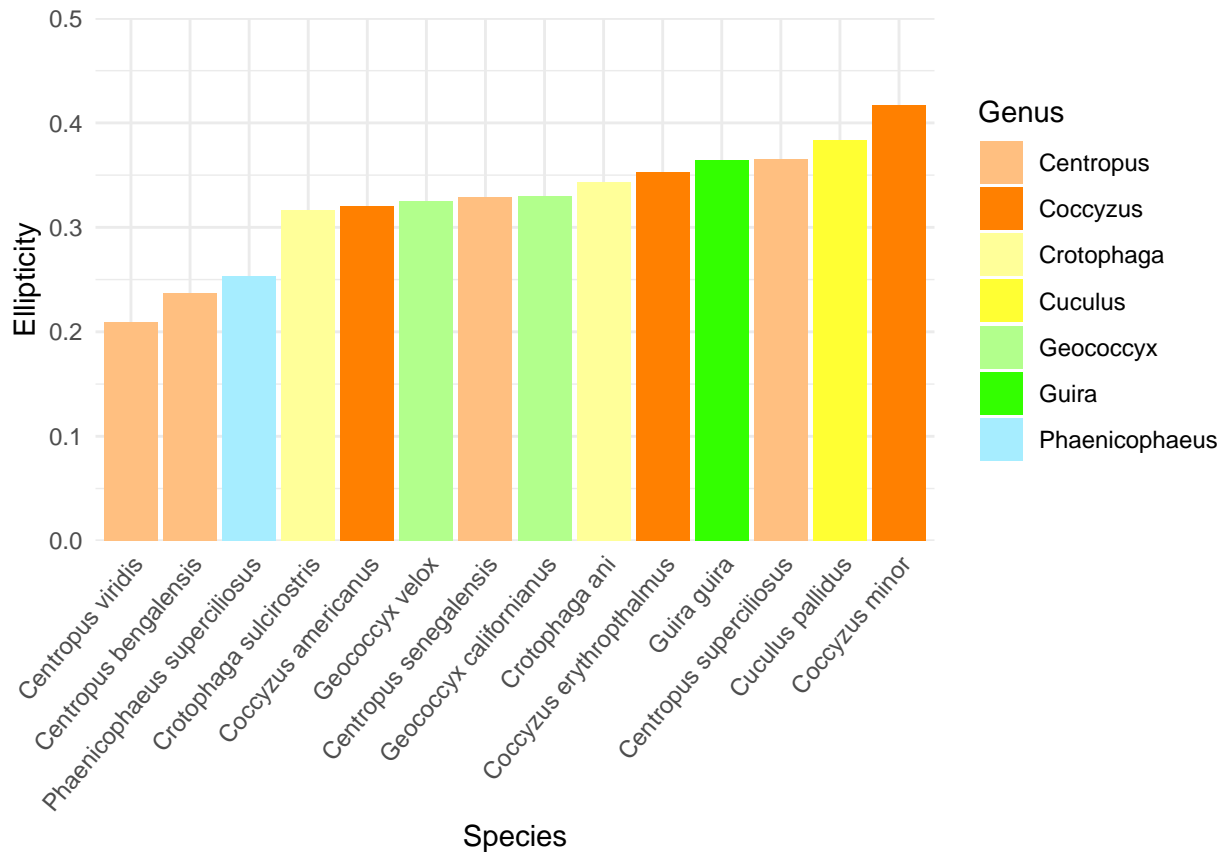
```
ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                         y=Ellipticity,
                                         fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,0.5,0.1),
                    limits=c(0,0.5),
                    expand=c(0,0)) +
  scale_fill_manual(values=dichromat(c("goldenrod","lawngreen","firebrick",
                                       "darksalmon","dodgerblue","darkviolet",
                                       "darkslateblue"),
                                   "protan")) +
  labs(x="Species") +
  theme_minimal() +
  theme(axis.text.x=element_text(angle=50, hjust=1))
```



dichromat also comes with some predefined color schemes in the `colorschemes` object. Here, we use `colorschemes$Categorical.12` because we're using categorical data, and subset the first 7 elements for our 7 levels:

```
ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                         y=Ellipticity,
                                         fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,0.5,0.1),
                    limits=c(0,0.5),
                    expand=c(0,0)) +
  scale_fill_manual(values=dichromat::colorschemes$Categorical.12[1:7]) +
```

```
labs(x="Species") +
theme_minimal() +
theme(axis.text.x=element_text(angle=50, hjust=1))
```



I find this one doesn't work great for me personally.

viridis

```
install.packages("viridis")
```

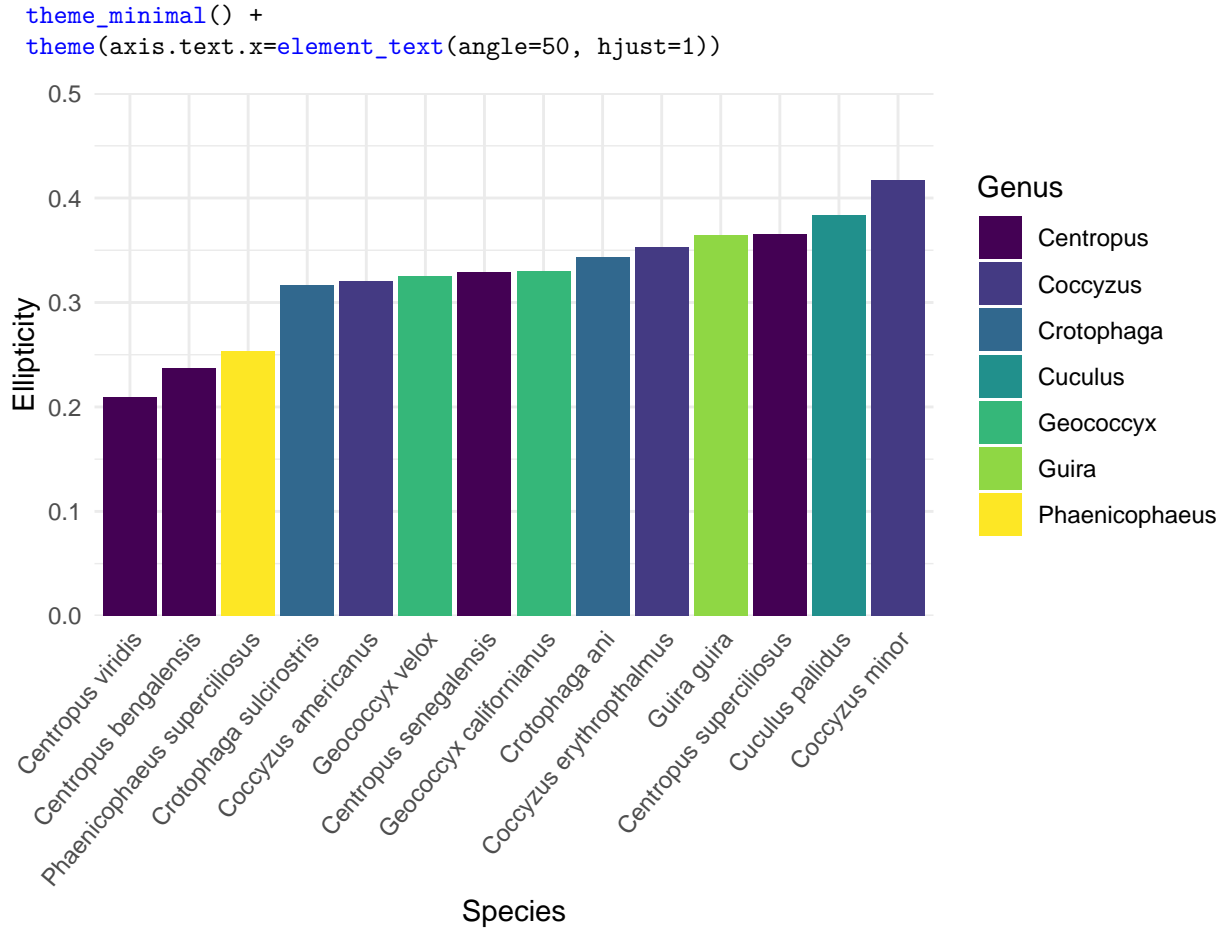
```
library(viridis)
```

```
## Warning: package 'viridis' was built under R version 3.5.3
```

```
## Loading required package: viridisLite
```

The `viridis` package has a number of color-blind friendly palettes, listed under their own functions that end with `_d` (for discrete) or `_c` (for continuous). The default is "viridis":

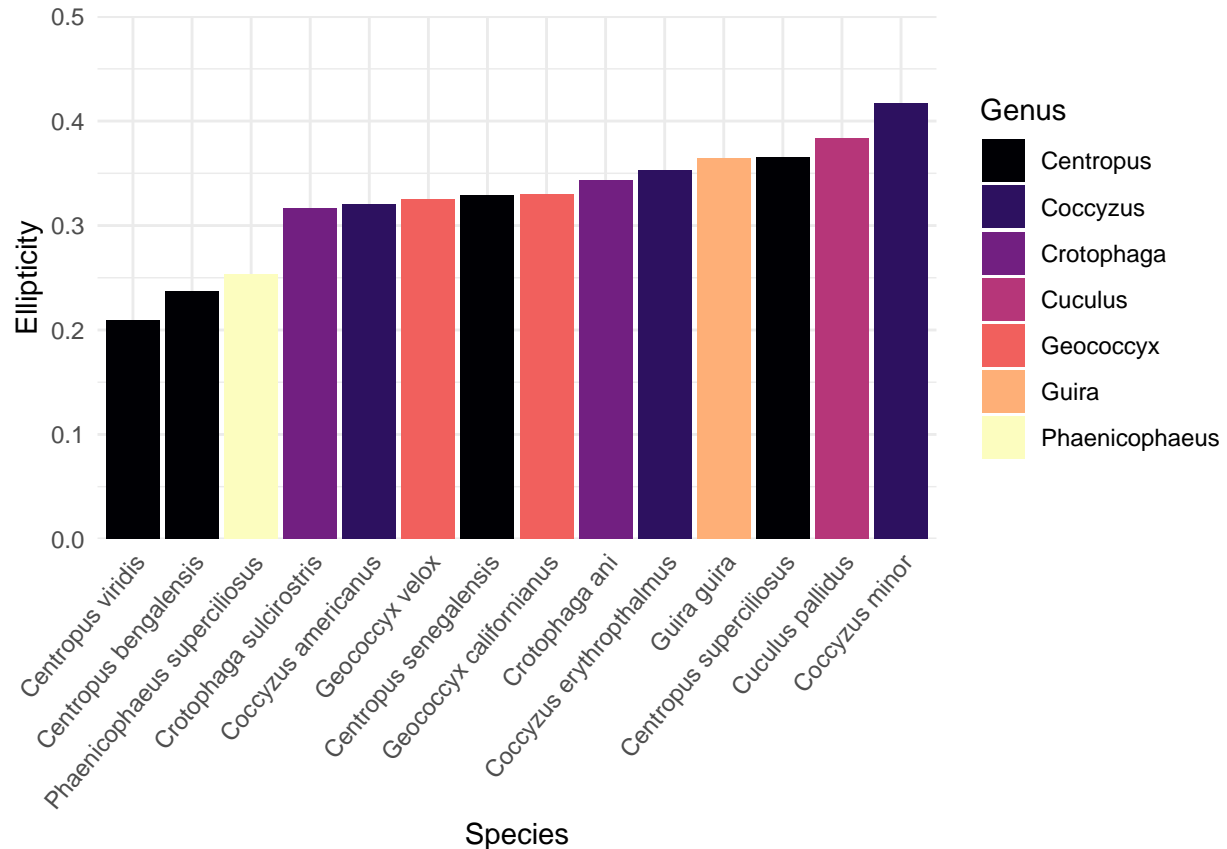
```
ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                     y=Ellipticity,
                                     fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,0.5,0.1),
                    limits=c(0,0.5),
                    expand=c(0,0)) +
  scale_fill_viridis_d() +
  labs(x="Species") +
```



Still not great for me, with all the dark blues/purples.

But there are others, such as "magma":

```
ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
y=Ellipticity,
fill=Genus)) +
geom_bar(stat="identity") +
scale_y_continuous(breaks=seq(0,0.5,0.1),
limits=c(0,0.5),
expand=c(0,0)) +
scale_fill_viridis_d(option="magma") +
labs(x="Species") +
theme_minimal() +
theme(axis.text.x=element_text(angle=50, hjust=1))
```



Again, not my preference, but it's there and could work.

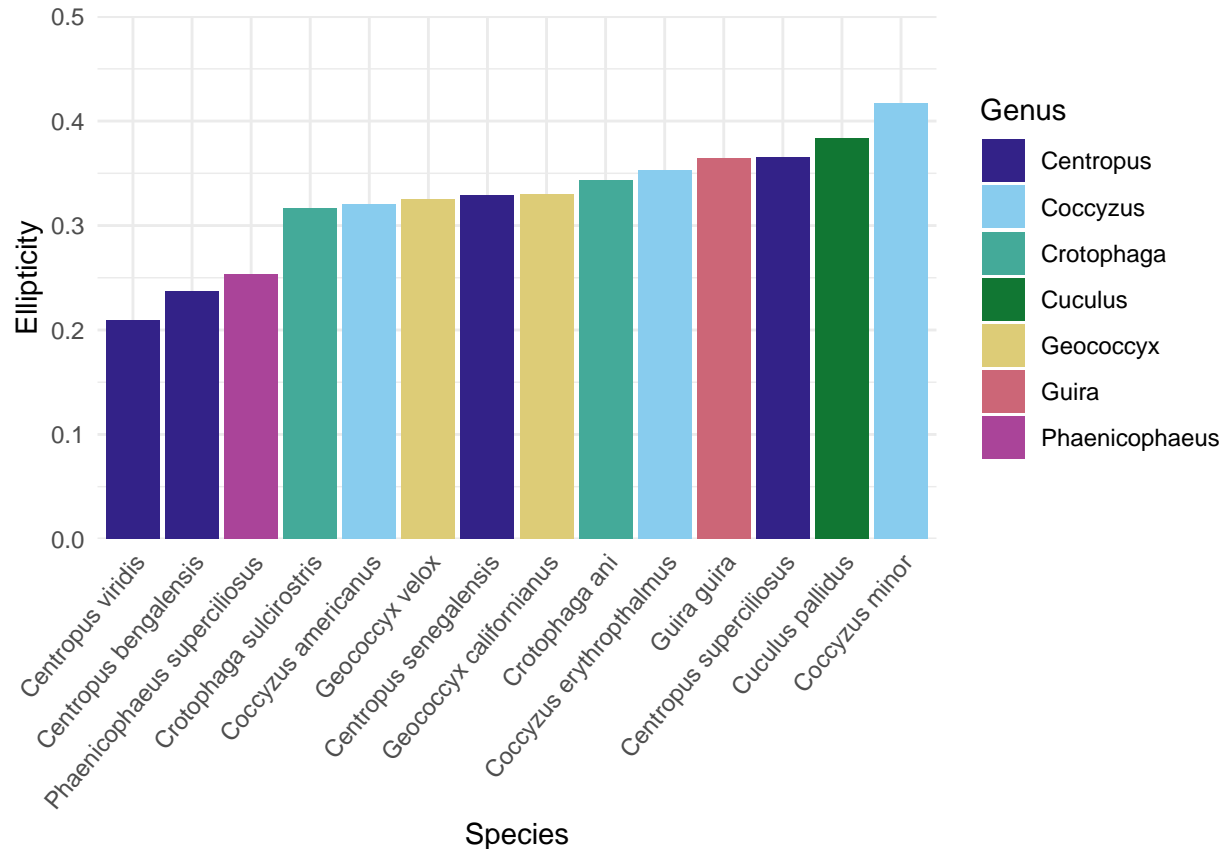
### Paul Tol

Finally, my absolute favorite:

Tol, P. (2018). Colour schemes. SRON Netherlands Institute for Space Research. Available: <https://personal.sron.nl/~pault/data/colourschemes.pdf>

The older version of his qualitative color scheme (which I actually like better than the new one) is available in the `ggthemes` package, which we already have loaded. Weirdly, you have to specify the number of levels in parentheses *after* the function, rather than as an argument in the function.

```
ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                     y=Ellipticity,
                                     fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,0.5,0.1),
                    limits=c(0,0.5),
                    expand=c(0,0)) +
  scale_fill_manual(values=ggthemes::ptol_pal()(7)) +
  labs(x="Species") +
  theme_minimal() +
  theme(axis.text.x=element_text(angle=50, hjust=1))
```



A variety of other color schemes are available in the document linked above which, in addition to being color-blind friendly, are also distinct in grayscale.

## Saving plots

`ggplot()`

First, assign your plot to an object:

```
plotCuckoo = ggplot(data=eggCuckoo, mapping=aes(x=reorder(Species, Ellipticity),
                                                  y=Ellipticity,
                                                  fill=Genus)) +
  geom_bar(stat="identity") +
  scale_y_continuous(breaks=seq(0,0.5,0.1),
                    limits=c(0,0.5),
                    expand=c(0,0)) +
  scale_fill_manual(values=ggthemes::ptol_pal()(7)) +
  labs(title="Egg ellipticity in Cuculiformes spp.", x="Species",
       caption="Data from Stoddard et al. (2017)") +
  theme_minimal() +
  theme(axis.text.x=element_text(angle=50, hjust=1))
```

Then give it as an argument to the `ggsave()` function:

**JPG**



```
ggsave("./figs/plotCuckoo.jpg", plotCuckoo, width=8, height=6, units="in",  
        dpi=300)
```

### PDF

```
ggsave("./figs/plotCuckoo.pdf", plotCuckoo, width=8, height=6, units="in")
```

### base

These functions let you save any kind of graphic, including `ggplots`. You have to use the graphic function first to open a “graphic device” (where the plot is sent), then run the code to produce the plot, then close the connection to the device with `dev.off()`.

### JPG

```
jpeg("./figs/plotCuckoo2.jpg", width=8, height=6, units="in",  
      res=300)  
plotCuckoo  
dev.off()
```

### PDF

```
pdf("./figs/plotCuckoo2.pdf", width=8, height=6)  
plotCuckoo  
dev.off()
```

(pdf / Rmd)